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RYANODYL 3-(PYRIDINE-3-CARBOXYLATE): A NOVEL RYANOID FROM RYANIA INSECTICIDE

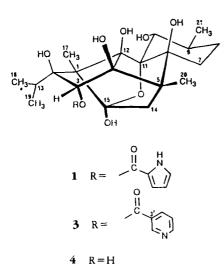
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ABSTRACT.—Ryanodyl 3-(pyridine-3-carboxylate) [3] was isolated as a minor component from the wet $CHCl_3$ extract of *Ryania* insecticide, and its structure was assigned by chemical and spectroscopic methods. This compound is essentially inactive compared with ryanodine for insecticidal activity against *Musca domestica* adults and *Tribolium castaneum* larvae, for toxicity to mice, and for competition with [³H]ryanodine at the Ca²⁺-ryanodine receptor complex of skeletal muscle.

Ryania insecticide is the powdered stemwood of Ryania speciosa Vahl. (Flacourtiaceae), a small tree or shrub growing extensively in South America and noted for its pest control properties (1). The major insecticidal and toxic constituents are ryanodine [1] (2,3) and didehydroryanodine [2] (4), which have attracted much attention as natural but expensive insecticides and for their action at the Ca^{2+} -ryanodine receptor complex of muscle (5). Variations in the extractives of different batches of commercial Ryania are known to occur, and these appear to contain additional minor constituents that also warrant attention from both of these points of view (6,7). We therefore extracted a quantity of R. speciosa powder using published methods (2,7) and by a combination of chromatographic procedures have isolated a new crystalline ryanoid **3**.

The ¹H- and ¹³C-nmr spectra of 3closely corresponded to those of $\mathbf{1}$ (8–10) with the exception of the aromatic resonances. This suggested that the alcohol portion, ryanodol [4], of 1 was retained but with a different esterifying group. More specifically, the signals characteristic of the pyrrole moiety were absent and instead, resonances were present whose ¹³C shifts and ¹H shifts and couplings were readily assignable to a pyridine-3-carboxylate. The hrms of 3requires C₂₆H₃₅NO₉ and is consistent with this proposal. The placement of the ester functionality at C-3 is corroborated by comparison of the chemical shift for the H-3 of **3** (δ 5.86) with that of **1** (δ 5.35). To verify the structure of 3, a



 $HO CH_{3} HO CH_{3} CH_{3} HO CH_{3} CH_{3} HO CH_{3} CH$

small sample was hydrolyzed using aqueous base. Following workup, ryanodol and pyridine-3-carboxylic acid were isolated and structurally confirmed by direct comparisons with authentic standards.

The isolation of the new ester 3 of ryanodol [4] is particularly interesting in relation to biosynthetic considerations, since esterification at this position cannot be effected under normal conditions; indeed, the total synthesis of ryanodine had to be curtailed to that of ryanodol (11). Because the introduction of the pyrrole ester moiety in ryanodine probably occurs at a late stage in the biosynthesis, an alternative to direct esterification at the 3 position could perhaps involve esterification at the contiguous and relatively accessible 4-hydroxyl with later acyl migration.

In view of the interest in Ryania as a natural insecticide, the new ester 3 was tested for toxicity to Musca domestica L. adults, Tribolium castaneum (Herbst) larvae, and mice, and for displacement of $[^{3}H]$ ryanodine from the Ca²⁺-ryanodine receptor complex. In each case, the level of activity was much less than that of ryanodine (8), and this compound is therefore considered to contribute neither to the insecticidal activity nor to the mammalian toxicity of the commercial mixture. Spatial considerations alone are unlikely to account for the profound effect that replacing the pyrrole ring by the pyridine ring has on the biological activity. Other differences include the electron-rich nature of the pyrrole and its ability to provide a hydrogen bond.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.— Hplc separation involved a Beckman 344-2 pump gradient system using a 10 mm \times 250 mm Ultrasphere-ODS C-18 reversed-phase Si column and MeOH/H₂O mixture at 4 ml/min with detection at 265 nm. ¹H- and ¹³C-nmr spectra were recorded with a Bruker WM-300 spectrometer; the solvent peaks were used as internal references. Hydroxyl peaks were confirmed by exchange with D₂O. Mass spectra were obtained using a Kratos MS-50 instrument. Radial chromatography was carried out using a model 8924 Chromatotron supplied by Harrison Research, Palo Alto, California, with one or two mm plates coated with Si gel 60 PF₂₅₄ (Merck). Workup solutions were dried with anhydrous Na_2SO_4 and evaporated with a rotary evaporator.

ISOLATION OF RYANOIDS.—Powdered R. speciusa wood supplied by Agrisystems International, Wind Gap, Pennsylvania, was extracted using the wet CHCl₃ procedure (2,7) modified by using a larger quantity of water (800 ml/500 g) for more efficient extraction. The extractives were partitioned into H2O and the aqueous phase was extracted with EtOAc. The EtOAc extract was separated from polar impurities by rapid filtration through Si gel in CHCl₃/MeOH (6:1) with 2% aqueous MeNH₂. The eluted material, in batches (0.4 g), was separated using CHCl₃-MeOH (20:6 to 6:1) with 2% aqueous MeNH₂ on a Chromatotron plate (2 mm Si gel). Ryanodyl 3-(pyridine-3-carboxylate) was present in a mixed fraction and was separated by hplc. The yield was ca. 50 mg/kg relative to the Ryania powder. Crystallization from Me₂CO/hexane gave 3 as prisms: mp 200-205°; ¹H nmr (Me₂CO- d_6) δ 9.21 (dd, 2.1, 0.9 Hz; H-2'), 8.85 (dd, 1.8, 4.5 Hz, H-6'), 8.41 (ddd, 1.8, 2.1, 7.8 Hz; H-4'), 7.61 (ddd, 1.0, 4.9, 7.8 Hz, H-5'), 5.86 (s, H-3), 3.85 (m, H-10), 2.60 and 2.08 (AB, 13.8, H₂-14), 2.35 (m, H-13), 1.82 (m, H-9), 2.12 (m, H_{ax} -7), 1.58 (m, H_{eq} -7), 1.50 (m, H_{ax} -8), 1.26 (m, H_{eg} -8), 1.47 (s, H_3 -17), 1.12 (d, 6.6 Hz, H3-21), 0.88 and 0.78 (each d, 6.5 and 6.6 Hz, H_3 -18 and H_3 -19); ¹³C nmr (Me₂CO-d₆) δ 164.7 (C=O), 154.6 (C-6'), 151.1 (C-2'), 137.8 (C-4'), 126.6 (C-3'), 124.7 (C-5'), 65.3 (C-1), 83.4 (C-2), 92.2 (C-3), 91.7 (C-4), 49.1 (C-5), 85.9 (C-6), 26.7 (C-7), 28.9 (C-8), 34.8 (C-9), 72.1 (C-10), 86.9 (C-11), 96.3 (C-12), 30.4 (C-13), 41.8 (C-14), 102.8 (C-15), 10.1 (C-17), 18.7 and 18.6 (C-18, C-19), 12.5 (C-20), 19.3 (C-21). Protonated carbons were assigned by the INVCOR technique (12).

HYDROLYSIS OF **3**.—The ester (21 mg) was hydrolyzed by refluxing for 5 h in 5% KOH/ MeOH. After addition of aqueous HCl to pH 4, the solution was evaporated and KCl separated with MeOH. The organic residue was filtered through Si gel in CHCl₃-MeOH (9:1) with 2% aqueous MeNH₂ to give ryanodol (10 mg), identified by R_f and ¹H-nmr spectrum. Later chromatographic fractions contained pyridine-3carboxylic acid (3 mg) identified by tlc and ¹H nmr (D₂O).

BIOLOGICAL TESTING.—Compounds 1 and 3were compared for toxicity to housefly adults (M. *domestica*) treated by injection following topical application of the synergist piperonyl butoxide, for toxicity to mice treated intraperitoneally, and for inhibition of [³H]ryanodine binding in a rabbit skeletal muscle preparation of the Ca²⁺ryanodine receptor complex (8). An additional test involved the inhibition of growth and development of first instar flour beetle larvae (*T. castaneum*) by dietary incorporation (13). In all of the above tests the new ester was essentially inactive compared with ryanodine.

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